

0960-894X(95)00079-8

## INHIBITION OF MATRIX METALLOPROTEINASES BY N-CARBOXYALKYL PEPTIDES CONTAINING EXTENDED ALKYL RESIDUES AT P1'

Craig K. Esser, \*a Ihor E. Kopka, a Philippe L. Durette, a Richard K. Harrison, b.c Lisa M. Niedzwiecki, b Maria Izquierdo-Martin, Ross L. Stein, b,d William K. Hagmanna

Departments of Medicinal Chemical Research and Enzymology<sup>b</sup>
Merck Research Laboratories
P.O. Box 2000, Rahway, New Jersey 07065-0900

Abstract: A series of N-carboxyalkyl peptides were prepared to test their inhibitory activity against human stromelysin (MMP-3), collagenase (MMP-1), and gelatinase A (MMP-2). Linear alkyl and  $\omega$ -aminoalkyl residues were employed as replacements for a phenethyl group yielding inhibitors with *in vitro* activities comparable to their corresponding aromatic analogs.

The matrix metalloproteinases (MMP's) are a family of zinc-containing, calcium dependent mammalian proteinases that are capable of degrading the extracellular matrix of connective tissues and basement membranes. These enzymes have been implicated in a variety of biological processes, including degradative diseases such as rheumatoid and osteoarthritis. Earlier work by our group established a structure-activity relationship (SAR) for the inhibition of MMP's by N-carboxyalkyl peptides. This SAR revealed a deep, hydrophobic pocket at the  $S_1$ ' subsite of stromelysin, a conclusion that has since been confirmed by NMR structure determination. The most potent stromelysin inhibitor to emerge from that effort was [N-1(R)-carboxyethyl)- $\alpha$ -(S)-(phenylethyl)]glycine-(S)-leucine, N-phenylamide, 1, with a  $K_i = 0.47 \mu M$ .

In 1990, Shirota and coworkers demonstrated that a long-chain, ω-aminoalkyl group could be used as a substitute for the phenethyl group in the angiotensin converting enzyme (ACE) inhibitor enalaprilat.<sup>5</sup> Since our lead compound 1 for stromelysin also contained the phenethyl side-chain, we explored a similar replacement in our molecule in an effort to improve its activities against MMP's and selectivity for stromelysin. To that end, we have prepared a series of analogs containing long-chain alkyl and ω-aminoalkyl groups at the P<sub>1</sub>' position and measured their activities against human stromelysin, collagenase, and gelatinase A.

<sup>&</sup>lt;sup>c</sup> Present address: 3D Pharmaceuticals 3700 Market Street, Philadelphia, PA 19104

d Present address: Mycogenics, Inc. 61 Moulton Road, Cambridge, MA 02139

## Chemistry

The synthesis of the inhibitors generally followed procedures as described in reference 3. The unnatural amino acids incorporated at the  $P_1$ ' position were prepared by stereoselective azide transfer methods described by Evans et al.<sup>6</sup> Scheme 1 illustrates the synthesis of [N-1(R)-carboxyethyl)- $\alpha$ -(S)-(9-aminononyl)]glycine-(S)-leucine, N-phenylamide as an example of our typical procedure. Starting with commercially available 11-aminoundecanoic acid 2a, the amino functionality was protected as its benzyl carbamate (Cbz) to furnish 3a. Stereoselective introduction of azide to the oxazolidinone 4a, followed by hydrolysis of the chiral auxiliary afforded the chiral  $\alpha$ -azido acid 6a in moderate yield. Coupling of the  $\alpha$ -azido acid 6a to amino acid anilides (L-leucinanilide, in this case) was accomplished using standard EDC/HOBt conditions.<sup>7</sup> Reduction of the azide in the presence of the Cbz group proceeded smoothly with SnCl<sub>2</sub>8 to give the dipeptide 8a. Displacement of the triflate<sup>9</sup> derived from benzyl (S)-lactate with the dipeptide 8a, followed by catalytic hydrogenation of the benzyl ester afforded the N-carboxyalkyl peptides listed in Table 1. Analogous procedures were followed for alkyl carboxylates 2b and 2c.

a) benzyl chloroformate, aq. NaHCO<sub>3</sub>, THF, 0°-25°; b) trimethylacetyl chloride, EtyN, THF, 0°, (S)-(-)-4-benzyl-2-oxazolidinone, n-BuLi, -78°; c) KN(SiMe<sub>3</sub>)<sub>2</sub>, Trisyl-N<sub>3</sub>, -78°, AcOH; d) LiOH, H<sub>2</sub>O<sub>2</sub>, THF, H<sub>2</sub>O; e) (L)-LeuNHPh or diCbz-(L)-ArgNHPh, HOBt, EDC, THF, 25°; f) SnCl<sub>2</sub> MeOH, 25°; g) benzyl (S)-lactate, Tf<sub>2</sub>O, 2,6-lutidine, Et(<sup>1</sup>Pr)<sub>2</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0-25°; h) H<sub>2</sub>, Pd(OH)<sub>2</sub>/C, MeOH.

## Results and Discussion

Compounds of the general structure shown below were evaluated *in vitro* against human fibroblast stromelysin (MMP-3), human fibroblast collagenase (MMP-1), and human gelatinase A (MMP-2). The data in Table 1 indicate that stromelysin and gelatinase A can accommodate extended hydrocarbon and  $\omega$ -amino hydrocarbon chains in their S<sub>1</sub>' pockets, while collagenase will not. Activities against stromelysin and gelatinase A were found to be comparable to those of the corresponding inhibitors containing a phenethyl group at the P<sub>1</sub>' position.<sup>3</sup> Similar results have been reported with hydroxamic acid inhibitors of MMP's with long-chain alkyl groups at P<sub>1</sub>'.<sup>10</sup>

In conclusion, we have demonstrated that linear alkyl chains can be substituted for a hydrophobic phenethyl group at P<sub>1</sub>' in this class of MMP inhibitors without adversely affecting the potency against stromelysin and gelatinase A. Subsequent structural determination of stromelysin has revealed a deep, hydrophobic pocket at S<sub>1</sub>' that extends completely through the enzyme that could easily accommodate these extended alkyl groups.<sup>4</sup> The X-ray crystal structure of human fibroblast collagenase indicates a much smaller S<sub>1</sub>' specificity pocket for this enzyme compared to stromelysin.<sup>11</sup> The loss of activity versus fibroblast collagenase with compounds 10-14 can be explained by the inability of this enzyme to fit a long alkyl chain in its much shallower S<sub>1</sub>' pocket.

Table 1. Inhibition of Human Stromelysin, Collagenase, and Gelatinase A by N-Carboxyalkyl Peptides with Long Chain Alkyl and Aminoalkyl Substituents at P1'

Compound Number	R <sub>1</sub> (P <sub>1</sub> ')	R <sub>2</sub> (P <sub>2</sub> ')	Stromelysin K <sub>i</sub> , µM (± S.E.)	Collagenase K <sub>i</sub> , µM (± S.E.)	Gelatinase A $K_i$ , $\mu M$ ( $\pm$ S.E.)
1	(CH <sub>2</sub> ) <sub>2</sub> Ph	i-C <sub>4</sub> H <sub>9</sub>	0.47 (0.08)	0.76 (0.22)	0.20 (0.04)
10	n-C9H <sub>18</sub> NH <sub>2</sub>	i-C <sub>4</sub> H <sub>9</sub>	0.24 (0.04)	> 10	0.50 (0.04)
11	n-C <sub>8</sub> H <sub>17</sub>	i-C <sub>4</sub> H <sub>9</sub>	0.57 (0.05)	>10	0.34 (0.05)
12	n-C8H17	(CH <sub>2</sub> ) <sub>3</sub> NHC(NH)NH <sub>2</sub>	1.60 (0.10)	>10	0.12 (0.03)
13	n-C9H <sub>18</sub> NH <sub>2</sub>	(CH <sub>2</sub> ) <sub>3</sub> NHC(NH)NH <sub>2</sub>	0.37 (0.04)	>10	0.11 (0.01)
14	n-C <sub>10</sub> H <sub>21</sub>	i-C <sub>4</sub> H <sub>9</sub>	0.85 (0.06)	>10	0.56 (0.11)

Stromelysin, collagenase, and gelatinase A assays were all performed at pH = 7.5 and 25° C according to the procedures detailed in Reference 3.

Acknowledgment: We thank Drs. C.G. Caldwell, K.T. Chapman, and V.L. Moore for their helpful suggestions and discussions.

## References and Notes

- a) Murphy, G. J. P.; Murphy, G.; Reynolds, J.J. FEBS Lett. 1991, 289, 4. b) Edonard, H.; Grimaud, J.-A. Cell. Molec. Biol. 1990, 36, 131. c) Matrisian, L. M. Trends Genet. 1990, 6, 121. d) Matrisian, L. M. Bioessays 1992, 14, 455.
- a) Dean, D. D.; Martel-Pelletier, J.; Pelletier, J.-P.; Howell, D. S.; Woessner, J. F. J. Clin. Invest. 1989, 84, 678. b) Hasty, K. A.; Reife, R. A.; Kang, A. H.; Stuart, J. M. Arthr. Rheum. 1990, 33, 388. c) Okada, Y.; Shinmei, M.; Tanaka, O.; Naka, K.; Kimura, A.; Nakanishi, I.; Bayliss, M. T.; Iwata, K.; Nagase, H. Lab. Invest. 1992, 66, 680. d) Walakovits, L. A.; Bhardwaj, N.; Gallick, G. S.; Lark, M. W. Arthr. Rheum. 1992, 35, 35.
- Chapman, K. T.; Kopka, I. E.; Durette, P. L.; Esser, C. K.; Lanza, T. J.; Izquierdo-Martin, M.; Niedzwiecki, L.; Chang, B.; Harrison, R. K.; Kuo, D. W.; Lin, T.-Y.; Stein, R. L.; Hagmann, W. K. J. Med. Chem. 1993, 36, 4293.
- Gooley, P. R.; O'Connell, J. F.; Marcy, A. I.; Cuca, G. C.; Salowe, S. P.; Bush, B. L.; Hermes, J. D.;
   Esser, C. K.; Hagmann, W. K.; Springer, J. P.; Johnson, B. A. Structural Biology 1994, 1, 111.
- 5. Shirota, M.; Kajiwara, Y.; Iijima, M.; Kitabatake, K. Drug Res. 1990, 40, 515.
- 6. Evans, D. A.; Bitton, T. C.; Ellman, J. A.; Dorow, R. L. J. Am. Chem., Soc. 1990, 112, 4011.
- 7. Bodanszky, M.; Bodanszky, A.; *The Practice of Peptide Synthesis*; Springer-Verlag: Berlin, Germany, 1984.
- 8. Maiti, S. N.; Singh, M. P.; Micetich, R. G. Tetrahedron Lett. 1986, 27, 1423.
- Attwood, M. R.; Hassall, A. K.; Krohn, A.; Lawton, G.; Redshaw, S. J. Chem. Soc., Perkin Trans. I 1986, 1011.
- Brown, P. A. unpublished results; Broadhurst, M. J.; Brown, P. A.; Johnson, W. H.; Lawton, G. Eur. Pat. Appl. #0575844A2, 1993.
- a) Lovejoy, B.; Cleasby, A.; Hassell, A. M.; Longley, K.; Luther, M. A.; Weigl, D.; McGeehan, G.; McElroy, A. B.; Drewry, D.; Lambert, M. H.; Jordan, S. R. Science 1994, 263, 375. b) Borkakoti, N.; Winkler, F. K.; Williams, D. H.; Arcy, A. D.; Broadhurst, M. J.; Brown, P. A.; Johnson, W. H.; Murray, E. J. Structural Biology 1994, 1, 106. c) Spurlino, J. C.; Smallwood, A. M.; Carlton, D. D.; Banks, T. M.; Vavra, K. J.; Johnson, J. S.; Cook, E. R.; Falvo, J.; Wahl, R. C.; Pulvino, T. A.; Wendoloski, J. J.; Smith, D. L. Proteins: Structure, Function, and Genetics 1994, 19, 98.

(Received in USA 14 September 1994; accepted 29 September 1994)